

Parasitism of *Clarias camerunensis* Lönnberg, 1895 (Siluriformes, Clariidae) by monogeneans in a dense tropical humid forest (Southern Cameroon-Africa) gives more arguments for fish quarantine in breeding using native species

Nack Jacques^{1,2}, Bassock Bayiha Etienne Didier³, Mbondo Jonathan Arme^{3,4}, and Bilong Bilong Charles Félix³

¹Faculty of Science, University of Douala, P.O Box 24157, Douala, Cameroon

²Institute of Fisheries and Aquatic Sciences, University of Douala, P.O Box 7236 Douala, Cameroon

³Laboratory of Parasitology and Ecology, Faculty of Science, University of Yaoundé I, P.O Box 812, Yaoundé, Cameroon

⁴Specialized Research Center for Marine Ecosystems, IRAD, P.O Box 219, Kribi, Cameroon

Copyright © 2020 ISSR Journals. This is an open access article distributed under the **Creative Commons Attribution License**, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

ABSTRACT: *Clarias camerunensis* is a potential catfish for farming in Cameroon. In order to assess the parasitism of its monogeneans as a function of season and standard length, a study was conducted from April 2017 to April 2018 in Lép Mōōga stream, of the Nyong river watershed (Southern Cameroon). 179 specimens of *C. camerunensis* were sampled by angling through 5 consecutive seasons. The Prevalence, density, abundance of adults and larvae of the main Monogenean species as well as the condition coefficient K and gonadosomatic index of the female *C. camerunensis* were calculated. The prevalence of the adult *Quadriacanthus* sp. remained equal to or greater than 85% during this study. Its abundance was low and did not show any clear profile pattern; however, its variation peaked during the rainy season. The gonadosomatic index (GSI) and condition factor (K) of female *C. camerunensis* evolved in parallel and showed that this fish lays three times a year, during the short rainy, the short dry and the long rainy seasons. The profile of *Quadriacanthus* sp. larvae showed that this monogenean breeds all year with two peaks during the short rainy season (when the female hosts lay and are weak fry also are in the environment), and in January when hosts are more concentrated in low water. The physiological condition of the fish gradually improved as they grew, when the parasitic density decreased. To limit monogenean outbreaks in farming, this work recommends that fish caught in the wild should be placed in quarantine and dewormed before being exploited.

KEYWORDS: parasitic worms, condition factor, *Clarias*, gonadosomatic index.

1 INTRODUCTION

The contribution of aquaculture and fisheries to the world economy in 2016 was 362 billion USD, of which 232 billion were obtained from aquaculture [23]. It is worth to note that from 2011 to 2016, the production of fishery resources fell from 92.2 million tons to 80 million tons [23]. As concerns African fish-farming in particular, a clear increase from 1.286 million tons in 2010 to 1.932 million tons in 2016 was observed [23]. In Cameroon, the consumer demand of fish is estimated at 400,000 tons [28]. In 2016 the total imports reached 151,858.5 tons of fish [28], while the national fish production was estimated at 180,000 tons, of which only 5,000 tons were obtained by fish farming [40]. This production by aquaculture clearly increased compared to that of 2012 estimated at 1000 tons [38]. Fish farming is therefore the main solution to increase fish production on the one hand; it also ensures employment for many africans on the other hand as it employed 304, 000 people in 2016 [23]. The breeding of better adapted native fish species is often recommended [2]; [61]. In Cameroon, several catfish species, among which *Clarias camerunensis* Lönnberg, 1895 whose length can exceed 46 cm are potential candidates for domestication and fish farming this is because of their good resistance to environmental conditions such as low oxygen concentration responsible

of death in fish ponds, temperature variations [54], and their rapid growth [53]; [22]. As a prelude to the domestication and breeding of catfish several studies have been undertaken in Africa [19] ; [56]; [2]; [60]. In the wild and even more in farming situations, catfish shelter many groups of parasites which can disturb their growth, their reproduction and consequently their yield [7], [30]. Many studies on parasites of siluriformes have been published in Africa [46]; [45]; [41]; [43]; [14]; [32]; [8]; [9]; [5]; [6]; [36]; [37]; [44]. In Cameroon, a few parasitological surveys carried out on catfish as well as on other freshwater fish have taken simultaneously into account the biology of the host and that of its parasites.

The aims of this work are therefore to: (1) follow the variations in the reproductive cycle of *Clarias camerunensis* and that of its xenocommunity of monogenean gill parasites, and (2) establish the relationships between these helminth loads, the condition factor and size of the fish.

2 MATERIAL AND METHODS

The fish were caught at night by angling from the Lép Mônga stream in the dense tropical humid forest of Ndjock Lipan (3 ° 41 '54''N;11 ° 5' 14 '' E) in the South centre plateau. Lép Mônga is a tributary of the Kéllé river, the longest tributary of the Nyong, the second longest river in Cameroon [47]. The climate in this ecosystem is a bimodal rainfall with 4 seasons: an irregular short dry season (SDS) from July to August, a long rainy season (LRS) from September to November, a long dry season (LDS) from December to mid-March and a short rainy season (SRS) from mid-March to June [52].

2.1 CONDUCT OF THE STUDY

Nine sampling campaigns were organized from April 2017 to April 2018: two per season, during the first four consecutive seasons, and one at the start of the second seasonal cycle. As soon as they were captured, fish were immediately immersed in a 10% formalin solution to fix and avoid the release of monogeneans due to an abundant post-mortem mucus secretion [11]. After the fish death, a small (1cm) abdominal incision was made to allow good conservation of its gonads [11]. Fish specimens were then transported to the laboratory.

2.2 MEASURED PARAMETERS

Before dissection, the standard length (SL: distance from the anterior end of the snout to the posterior end of the last vertebra of the fish [51]) was measured using a ruler graduated to the nearest millimeter. The mass of the carcass (Mc) of the fish i.e. stripped of its digestive tract and of the gonads (Mg) only for female individuals were also measured using an electronic Sartorius balance to the nearest hundredth gram.

2.3 RESEARCH, ASSEMBLY AND IDENTIFICATION OF MONOGENEAN GILL PARASITES

Gill arches were removed and placed in Petri dishes containing tap water. Monogeneans attached to the gill filaments were dislodged under a binocular WILD HEERBRUGG stereomicroscope, using a needle, then mounted between a slide and coverslip in a drop of eosin hematoxylin [11]. Parasite species identification was made based on sclerotized parts of the haptor and the copulatory organs after [15], [14] using the Leica DM 2500 microscope. The same preparations were used to count the different monogenean adults species and larvae.

2.4 CALCULATED PARAMETERS

The Prevalence (Pr) and abundance (A) are herein defined after [18], infracommunity and xenocommunity after [20]. The status of each parasite species was determined according to [59]: the species was stated frequent or common or even main or core (Pr > 50%), infrequent or secondary or intermediate ($10\% \leq Pr \leq 50\%$), and rare or satellite (Pr < 10%). The subsequent analysis concerned only the core species since in a community, satellite or rare species are little or no structuring [26]; [20]. The parasitism was categorized based on average helminthic load and according [12]. Therefore, the mean load (abundance) was described as very low ($A < 10$); low ($10 \leq A \leq 50$); average ($50 < A \leq 100$) and high ($A > 100$). The condition factor (K) was computed according to [58]; its formula is: $K = 10Mc / SL$; normally K increases regularly as the fish grows. The gonadosomatic index (GSI) indicates the relationship between the female gonads mass (Mg) and the body weight of a fish [58]; its formula is: $GSI = Mg / Mc - Mg$. In a fish population, the decrease of both K and GSI indicates the laying time [33]; [4]; [27]; [24]. The parasite density (D) is the ratio between the total number of monogeneans (Np) of the infracommunity and the host body weight : $D = Np / Mc$ [18].

2.5 STATISTICAL ANALYSIS

Comparison of prevalences was done using the Chi-square test (χ^2). The one-way analysis of variance (ANOVA) permitted to compare larval mean abundances, K and GSI during the sampling campaigns. For the follow up of the parasite density and the condition factor as a function of the fish standard length, a linear regression model was used. The software SPSS was used during all the analysis. Differences were considered as significant at $P < 0.05$.

3 RESULTS AND DISCUSSION

3.1 RESULTS

A total of 179 fish were sampled from April 2017 to April 2018, from Ndjock Lipan dense tropical forest: 33 fish in SRS, 42 in SDS, 24 in LRS 58 in LDS and 22 in SRS of the second cycle.

3.1.1 COMPOSITION OF THE MONOGENEAN XENOCOMMUNITY OF *CLARIAS CAMERUNENSIS*

A total of 4482 monogenean specimens were recorded from *Clarias camerunensis*. These worms belonged to four species: *Quadriacanthus* sp. ($n_1= 3938$: 87.86%), *Quadriacanthus teugelsi* Birgi, 1988 ($n_2= 36$: 0.80%), *Birgiellus kellensis* Bilong Bilong, Nack&Euzet, 2007 ($n_3= 402$: 8.97%) and *Gyrodactylus camerunensis* Nack, Bilong Bilong & Euzet, 2005 ($n_4= 106$: 2.37%). *Quadriacanthus* sp. was thus numerically predominant.

3.1.2 TEMPORAL VARIATION OF THE PREVALENCE AND ABUNDANCE OF *QUADRIACANTHUS* SP. (ADULTE STAGE)

The prevalence of the different monogenean species were 99.2%, 84.6% 64% and 42% for *Quadriacanthus* sp., *Birgiellus kellensis*, *Gyrodactylus camerunensis* and *Quadriacanthusteugelsi* respectively. In this enumeration order, their mean abundances were 22.8, 3.04, 4.29 and 0.20. Therefore, in the Ndjock Lipan ecosystem *Quadriacanthus* sp. was the main (core) species of the *C. camerunensis* monogenean xenocommunity; its prevalence remained very high $> 80\%$ (Fig. 1) and was relatively lower in April 2017, although these variations were not significant ($\chi^2 = 13.25$; $df = 8$; $P > 0.05$). The abundance of this ectohelminth was low over the study period (Fig. 2); it did not show any clear variation pattern despite a slight tendency to increase during the rainy seasons (May 2017 in SRS; October 2017 in LRS and April 2018 in SRS). However these fluctuations were not statistically significant ($F = 1.74$; $df = 8$; $P > 0.05$).

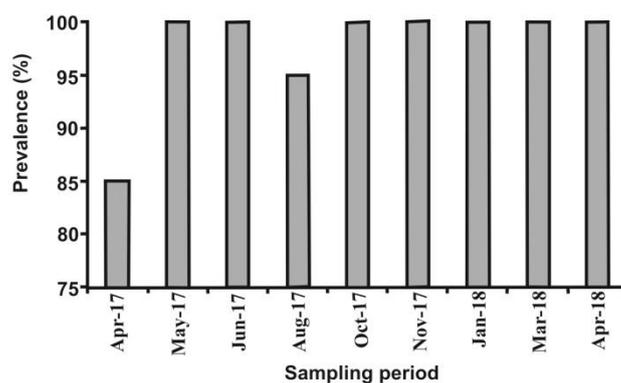


Fig. 1. Monthly prevalence of *Quadriacanthus* sp. during the sampling period

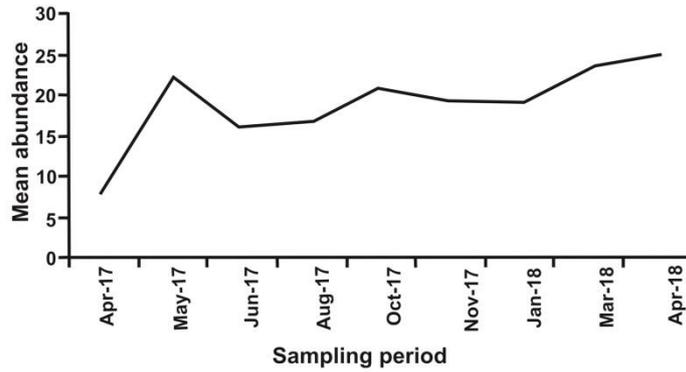


Fig. 2. Adult monthly mean abundance of *Quadriacanthus sp.* during the sampling period

3.1.3 TEMPORAL VARIATION OF THE ABUNDANCE OF *QUADRIACANTHUS SP.*(LARVAL STAGE)

Quadriacanthus sp. bred all year round in the dense tropical forest of Ndjock Lipan (Fig. 3). The larval abundance profile was a bimodal curve. The first peak appeared in May 2017 (SRS) and the second, more important, in January 2018 (LDS). The variations in larval mean abundance between the different sampling campaigns were significant ($F = 1.84$; $df = 8$; $P = 0.007$).

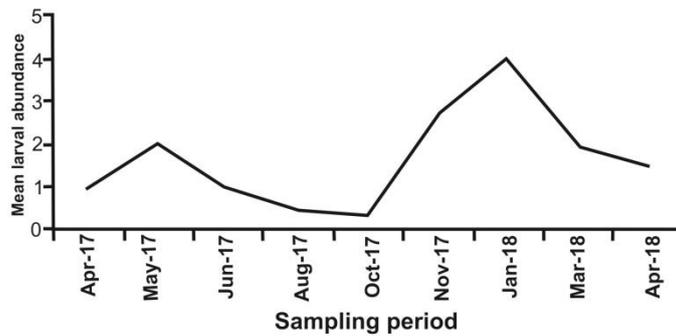


Fig. 3. Monthly mean larval abundance of *Quadriacanthus sp.* during the sampling period

3.1.4 TEMPORAL VARIATION OF THE FEMALE *CLARIAS CAMERUNENSIS* GONADOSOMATIC INDEX (GSI) AND CONDITION FACTOR (K)

The gonadosomatic index varied between the different sampling campaigns ($F = 1.92$; $df = 8$; $P = 0.003$). It showed two increase periods (June to August 2017 in SDS, November 2017 to March 2018 in LDS) and three decrease periods (April to June 2017 in SRS, August to November 2017: late SDS to early LRS; March to April 2018: early SRS) see figure 4. The condition factor (K) of female *C. camerunensis* varied significantly between the different sampling campaigns ($F = 2.00$; $df = 8$; $P = 0.001$, Fig. 4). It decreased from April to June 2017(SRS).It then increased somewhat in August to October (SDS to the first half of the LRS). In the second half of the LRS, it dropped. Another increasing phase was observed in the LDS followed by a fall from the second seasonal cycle (LDS to SRD 2018).

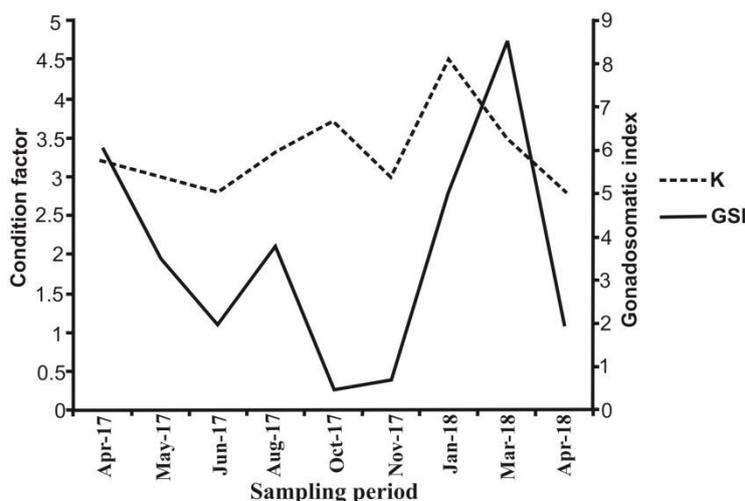


Fig. 4. Monthly gonadosomatic index GSI and condition factor K during the sampling period

3.1.5 RELATIONSHIP BETWEEN THE PARASITE DENSITY, THE CONDITION FACTOR AND THE HOST STANDARD LENGTH (REGARDLESS OF SEX)

The monogenean density decreased while the host standard length increased according to a linear model (Fig. 5: $D = -51 \times 10^4 LS + 1.38$; $r = -0.51$; $P = 0.0001$). As concerns the condition factor, values higher than 4.3 corresponded to those of males. K increased positively with the *C. camerunensis* standard length according to a linear model (see Fig. 6: $K = 35 \times 10^3 LS - 2.71$; $r = 0.94$; $P = 0.0001$).

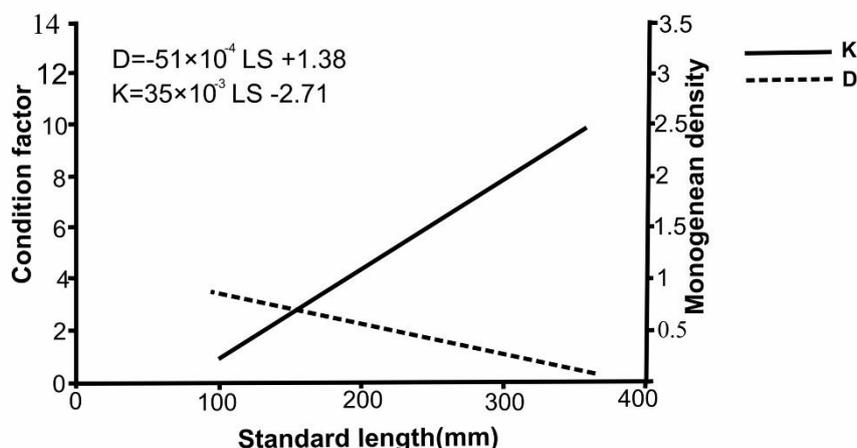


Fig. 5. Gonadosomatic index and condition factor as a function of standard length of *C. camerunensis* (regardless of sex)

In the Ndjock Lipan forest ecosystem, the monogenean density decreased when the condition factor of *Clarias camerunensis* increased according to a linear model ($D = -0.116K + 0.881$). The physiological condition of the host appeared negatively correlated ($r = -0.47$; $P = 0.004$) to the helminth load (density).

3.2 DISCUSSION

During the current study, no significant temporal variation in the abundance or prevalence of adults *Quadriacanthus* sp., the core species of this monogenean xenocommunity, was observed, although the abundance profile seemed to show 3 peaks during the rainy seasons. Moreover, the variation in abundance of *Quadriacanthus* sp. larvae indicated that this species bred throughout the year but mainly in two periods: May (SRS) and January (LDS). This result is close to that of [10] on *Gyrodactylus*

linstowi in Tajikistan (Central Asia) in a stream with constant temperature; these authors also didn't note any clear pattern in the parasitism as a function of season. Although with insignificant differences, our observation is similar to those by [46], [12], [13], [16], [3] and [39] respectively, for monogenean gill parasites of *Tilapia* sp. in Ghana *Chrysichthys nigrodigitatus* in Nigeria, *Hemichromis fasciatus* (now correctly renamed *H. elongatus* [51] and *Barbus martorelli* in Cameroon, *Sarotherodon melanotheron* and *Oreochromis niloticus* in Côte d'Ivoire, *Cyprinus carpio* in Algeria; these different monogenean communities showed outbreaks during the rainy seasons and spring. These results differ from those of [13] who did not find any model in the occurrence of monogenean larvae of *Barbus martorelli* in Cameroon. The constant prevalence and the low parasite abundance of *Quadriacanthus* sp. in the present study can be justified by the constancy of ambient and / or water temperature [10]; [46]. The decline of the gonadosomatic index (GSI) and condition factor (K) allows to identify the fish laying time [27]; [1]; [24]. Those of female *Clarias camerunensis* appeared seasonal, and cyclical and indicated that this catfish breeds three times a year during the short and long rainy seasons. Our results somewhat do not agree with those of [31] who pointed out that, in general the species of *Clarias* have only one laying period limited to a few months. However, they also claimed that, considering the bimodal distribution of their oocytes, these fish can breed all year round when conditions allow. In Nigeria [29] observed that *Parachanna obscura* bred most of the year (except in February, March and April) with two peaks in May and July. *Quadriacanthus* sp. larvae were more abundant in May 2017 and January 2018; it is thus suggested that this ectohelminth could have selected these periods because host fish are available. In the short rainy season, these larvae hatch when female hosts lay and their physiological condition and therefore their immune defenses are reduced; these weakened females, as well as the fry, are easy targets for the parasites. This is the reason why [55] argued that the ephemeral nature of the host as a habitat has forced parasites to find ways to colonize new individuals. They also indicated that the hatching of monogenean eggs is closely linked to environmental cycles and direct host stimuli. The relative outbreak of larvae in January in the Ndjock Lipan ecosystem can be explained by the availability of host individuals. In LDS the probable physicochemical modifications of the environment due to the shallow water causes an aggregation of the hosts which become easily accessible to infesting stages of parasites [49].

In this study, the condition factor (K) of *C. camerunensis* increased regularly as it grew, while the parasite density (D) decreased. Therefore, as the fish grows it acquires immunity limiting the parasite load. [34] and [35] also found a negative correlation between the total number of two gill monogenean parasites of *Piaractus mesopotamicus* and two others of *Arapaima gigas* respectively and the host condition factor K. Similarly, [50] observed in *Thymallus thymallus*, parasitized by *Gyrodactylus salaris* that the host gradually acquires an immune response limiting infestations. In contrast, [57] found no difference between the condition factors of the parasitized *Corydoras aeneus* individuals and those not parasitized. As concerns [25], they obtained a positive correlation between the abundance of four monogenean species and the condition factor of *Hoplias aff. malaboricus* in Brazil.

The parasite density decreased as a function of the standard length of *Clarias camerunensis*. [35] also observed a decrease in the number of monogeneans as a function of the body weight of *Arapaima gigas* but they didn't explain why. In the current work, the drop in the parasite density as a function of the standard length of *Clarias camerunensis* can be more explained by the fact that *Clarias* species have a rapid growth [17]; [53], since it was shown that the monogenean mean abundance was not host length dependant. Indeed, [48] and [21] respectively showed that in *Cyprinus carpio* and *Salmo gardneri*, the rapid multiplication of parasites is counteracted by the rapid growth of these fish limiting the damage.

4 CONCLUSION

This work, carried out in a dense tropical humid forest less anthropized, suggests that monogenean gill parasites of *Clarias camerunensis*, mainly *Quadriacanthus* sp. the core species of this xenocommunity, adopt strategies for a sustainable colonization of their host. This observation is illustrated by: a wide dispersal within the host population (high prevalence) and low abundance which do not reach a dangerous threshold throughout the year. Monogeneans reproduce when young fishes with weak immunity arrive in the environment, when the physiological condition of adult fish decreases and when all the fish population is more accessible because of increased concentration in shallow water during the dry season. To limit parasite outbreaks in farming, we suggest that the fish caught in the wild be placed in quarantine and undergo deworming before being exploited.

CONFLICT OF INTEREST

The authors declare no conflicts of interest.

ACKNOWLEDGEMENTS

- The study was funded by the special research allowances from the Ministry of higher education and internal allowances from University of Douala and University of Yaounde I.
- Thanks to Dr Clarisse Njua for technical advice assistance

REFERENCES

- [1] E. H. Alhassan, S. Abobi, and F. Boti, "The spawning pattern, length-weight relationship and condition factor of elephant fish, *Mormyrus rume* from the Bontanga reservoir, Ghana", *International Journal of Fisheries and Aquaculture Studies*, vol. 2, no. 2, pp. 109-114, 2014.
- [2] Y. L. Alla, M. Ouattara, M. C. Blé, and B. C. Atse, "Etude de la fécondité du silure Africain *Heterobranchus bidorsalis* (Geoffroy Saint-Hilaire, 1840) en conditions d'élevage". *Tropicicultura*, vol.28, no.1, pp. 31-36, 2010.
- [3] A. Allagua, S. Guerfi, N. Kaouach, C. Boualleg, I. Boucenna, C. Barour, A. Menasria, and M. Bensouilah, "L'infestation de *Cyprinus carpio* (cyprinidés) peuplant le barrage Fom el-Khanga (Souk ahras, Algérie) par les monogènes parasites", *Bulletin de la Société Zoologique de France*, vol.140, no.3, pp 217-232, 2015.
- [4] k. Amenzoui, N. Ferha, F. Tachinante, A. Yahyaoui, S. Kifani and A. K. Mesfioui, "analysis of the cycle of reproduction of *Sardina pilchardus* (Walbaum, 1792) off the moroccan atlantic coast", *Comptes Rendus Biologies*. vol. 329, pp.892-901, 2006.
- [5] D. N. D. Bahanak, J. Nack, A. R. Bitja Nyom, A. Pariselle and C. F. Bilong Bilong, "Quadriacanthus spp. (Monogenea, Dactylogyridea) from *Heterobranchus longifilis* (Teleostei, Clariidae) in the river Boumba (Congo basin: Cameroon) with the description of three new species". *Vie et milieu*., vol. 67. no. 2, pp 81-89, 2017.
- [6] D. N. D. Bahanak, J. Nack, A. Pariselle and C. F. Bilong Bilong, "Description of three new species of *Quadriacanthus* (Monogenea: Ancyrocephalidae) gill parasites of *Clarias submarginatus* (Siluriformes: Clariidae) from Lake Ossa (Littoral region, Cameroon)", *Zoologia*, vol. 33, no.4, pp.1-8, 2016.
- [7] Balarin, Hatton, Tilapia *A guide to their biology & culture in Africa*. University of Stirling, Scotland. 175p, 1979.
- [8] E. D. Bassock Bayiha, J. Nack, A. R. Bitja Nyom, A. Pariselle, and C. F. Bilong Bilong, "Description of three new species parasites from *Chrysichthys nigrodigitatus* and *Chrysichthys longidorsalis* (Siluriformes, Claroteidae) in the Sanaga river (Cameroon)", *Vie et Milieu*. vol. 67 no. 2, pp 65-73, 2017.
- [9] E. D. Bassock Bayiha, J. Nack, A. Pariselle and C. F. Bilong Bilong, "Two new species of gill parasites assigned to *Protoancylo-discoides* (Monogenea, Ancyrocephalidae) from *Chrysichthys* spp. (Siluriformes, Claroteidae) in River Sanaga (Cameroon)", *Zootaxa*. vol. 4170, no.1, pp 178-186, 2016.
- [10] V. Bauer and S. B. Karimov, "Pattern of parasitic infections of fishes to water body with constant temperature", *Journal of Fish Biologie*. vol. 36, pp 1-8, 1990.
- [11] Bilong Bilong, *Les monogènes parasites des poissons d'eau douce du Cameroun : Biodiversité et spécificité ; biologie des populations inféodées à Hemichromis fasciatus*. Thèse de Doctorat d'État, Université de Yaoundé I, 341 p, 1995
- [12] C. F. Bilong Bilong and T. Njiné, "Dynamique de populations de trois Monogènes parasites d'*Hemichromis fasciatus* (Peters) dans le lac municipal de Yaoundé et intérêt possible en pisciculture intensive", *Annales de la Faculté des Sciences de Yaoundé, Biologie-Biochimie. Série Sciences Naturelles et Vie*. vol. 34, pp 295-303, 1998.
- [13] C. F. Bilong Bilong and J. Tombi, "Temporal structure of a component community gill parasites of *Barbus martorelli* Roman, 1971 (Freshwater Cyprinidae) in the Centre province, Cameroon", *Cameroon Journal of Biological and Biochemical Sciences*, vol.13, pp 9-13, 2005.
- [14] C. F. Bilong Bilong, J. Nack and L. Euzet, "Monogènes de *Clarias* (Siluriformes, Clariidae) au Cameroun: II. Description de trois nouvelles espèces du genre *Birgiellus* n. gen. (Dactylogyridea, Ancyrocephalidae) dans le bassin du Nyong", *Parasite*, vol.14, pp 121-130., 2007.
- [15] E. Birgi, "Monogènes du genre *Quadriacanthus* Paperna, 1961, parasites branchiaux de deux Siluridae (Teleostei) *Clarias pachynema*, Boulenger, 1903, et *Clarias jaensis* Boulenger, 1909 au Sud-Cameroun (description de 4 espèces nouvelles)", *Annales de la Faculté des Sciences de Yaoundé, Biologie-Biochimie*, vol. 3, no. 5, pp 113-129, 1988.
- [16] K. G. Blahoua, V. N'douba, K. Tidiani and K. J. N'guessan, "Variations saisonnières des indices épidémiologiques de trois monogènes parasites de *Sarotherodon melanotheron* (Pisces : Cichlidae) dans le lac d'Ayamé (Côte d'Ivoire)", *Sciences & Nature*, vol.6, no.1, pp 39-47, 2009.
- [17] Bruton, M. N. *The breeding biology and early development of Clarias gariepinus* (Pisces : Clariidae) in Lac Sibaya, South Africa. With a review of breeding. In species of subgenus *Clarias* (*Clarias*). *Transactions of the Zoological Society of London*. vol.36 pp 1- 45, 1979.

- [18] A. O. Bush, K. D. Lafferty, J. M. Lotz, and A.W Shostak, "Parasitology meets ecology on its own terms", *The Journal of Parasitology*, vol. 83, pp 575-583, 1997.
- [19] A. Chikou, P.A. Laleye, V. Raemakers and V.J. Philippart, "Etude de l'âge et de la croissance chez *Clarias gariiepinus* (Pisces, Clariidae) dans le delta de l'Ouémé au Bénin (Afrique de l'Ouest)", *International Journal of Biological and Chemical Sciences*, vol. 2, no. 2, :pp 157-167, 2008.
- [20] Combes, *Interactions durables. Ecologie et évolution du parasitisme*. Masson. 524p. 1995.
- [21] R. Cussack, "Development of infections of *Gyrodactylus colemanensis* Mizelle and Kritsky, 1967 (Monogenea) and the effect on fry of *Salmo gairdneri* Richardson", *The Journal of Parasitology*, vol.72, no.5, pp 663-8, 1986.
- [22] D. Etaba Angoni, M. Tomedi Eyango, H. Djoko and J. Tchoumboué, "Performances de croissance du poisson-chat Africain *Clarias jaensis* Boulanger, 1909 (Pisces : Clariidae) en étangs fertilisés des fientes de poules et des lisiers de porcs", *International Journal of Innovation and Applied Studies*, vol.17, no.4, pp 1294-130, 2016.
- [23] FAO. La situation mondiale des pêches et de l'aquaculture 2018. Atteindre les objectifs de développement durable. Rome. Licence: CC BY-NC-SA 3.0 IGO : 1237p, 2018.
- [24] Z. Fatima Bouhali, B. Shahnaz lechekha, S. Iadaimia, Assia Bedairia, R. Amara and B. Abdallah Djebbar, "Reproduction et maturation des gonades de *Sardina pilchardus* dans le golfe d'Annaba (Nord-Est algérien)", *Cybium*, vol.39, no.2, pp 143-153, 2015.
- [25] R. J. Graça, A. P. L. Costa and R. M. Takemoto, "Ecological aspects of monogenea gill parasites (Platyhelminthes) from *Hoplias aff. malabaricus* (Bloch, 1794) (Pisces, Erythrinidae) in a neotropical floodplain," *Neotropical Helminthology*, vol.7, no.1, pp105-116, 2013.
- [26] J. C. Holmes, "The structure of helminth communities", *International Journal for Parasitology*. vol.17, pp 203–208, 1987.
- [27] L. Ibtissem, B. A. Mossadok and B. H. Oum Kalthoum, "Cycle de reproduction et maturité sexuelle de *Zosterisessor ophiocephalus* (Gobiidae) sur les côtes nord de la Tunisie (Lagune de Bizerte)", *Revue F. S. B.*, vol.10 pp 86-99, 2013.
- [28] INS. *Elevage et Pêche*. In Annuaire Statistique du Cameroun. *INS, Edition 2017*, 208-210p. 2017
- [29] I. A. Isangedighi, and O. E. Umoumoh, "Some aspects of the reproductive biology of african snakehead – *Parachanna obscura* in Itu-Cross River system," *Nigerian Journal of Agriculture, Food and Environment*, vol.7 no.4, pp 19-30, 2011.
- [30] Jackson, P. B. N. *Aquaculture in Africa*. In C. Lévêque, M. N. Bruton & G.W. Ssentongo (Eds), *Biologie et Ecologie des poissons d'eau douce africains*. l'ORSTOM., 508p. 1988.
- [31] Legendre, M., and Jalabert, B. *Physiologie de la reproduction*. In : Lévêque, C. (ed.), Bruton M. N. (ed.), Ssentongo G.W. (ed.), *Biologie et écologie des poissons d'eau douce africains* Paris : ORSTOM, vol.216, pp 153-175. 1988.
- [32] G. B. Lekeufack Folefack, and A. Fomena, "Structure et dynamique des infracommunautés de Myxosporidies parasites de *Ctenopoma petherici* Günther, 1864 (Anabantidae), *Clarias pachynema* Boulenger, 1903 (Clariidae) et *Hepsetus odoe* (BLOCH, 1794) (Hepsetidae) dans la rivière Sangé au Cameroun", *International Journal of Biological and Chemical Sciences*. vol.7, no.6, pp 2301-2316, 2013.
- [33] M. Lizama, and A. M. Ambrosio, "Condition factor in nine species of fish of the Characidae family in the upper paraná river floodplain, brazil", *Brazilian Journal of Biology*. vol. 62, no.1, pp 113-124, 2002.
- [34] M. Lizama, R. M. Takemoto, M. J. T. "Ranzani-Paiva, L. M. D. S. Ayroza and G.C. Pavanelli, Relação parasito hospedeiro em peixes de pisciculturas da região de Assis, Estado de São Paulo, Brasil", *Piaractus mesopotamicus* (Holmberg, 1887). *Acta Scientiarum-Biological*. Vol.29, no. 4,pp 437-445, 2007.
- [35] R. G. B., Marinho, M.Tavaresdias, M. K. R. Dias Grigório, L. R. Neves, E. T. O. Yoshioka, C. L. Boijink, and R. M. Takemoto, "Helminthes and protozoan of farmedpirarucu (*Arapaima gigas*) in eastern Amazon and host-parasite relationship", *Arquivo. Brasileiro de Medicina Veterinária e Zootecnia*, vol.65, no.4. pp 1192-1202, 2013.
- [36] J. A Mbondo, J. Nack, Bitja A. R. Nyom, A. Pariselle and C .F Bilong Bilong, "New species of *Synodontella* (Monogenea, Ancyrocephalidae) gill parasites of two *Synodontis* spp. (Pisces, Mochokidae) from the Boumba River (Congo Basin, East Cameroon)", *Parasite*. vol. 26, no. 37, : 1-8, 2019.
- [37] J. A. Mbondo, J. Nack, A. Pariselle and C. F Bilong Bilong "The diversity of monogenean gill parasites of two *Synodontis* species (Siluriformes, Mochokidae) with the description of two new species assigned to *Synodontella*", *Vie et Milieu*, vol. 67, :pp 75-80, 2017.
- [38] MINEPIA, Recensement des fermes piscicoles dans les zones à fort potentiel au Cameroun (Centre, Est, Ouest, Nord-Ouest et Sud). Rapport principal. 32p, 2013.
- [39] O. E. Mircea, N. M. Cioran, and S. Hagi Ionesco, "Eco-biological aspects of Monogeneans' gill parasites of *Oreochromis niloticus* in Lake Ayamé I, Côte d'Ivoire", *International Journal of Agriculture and Agricultural Sciences*, vol. 4, no. 1, pp 059-071, 2017.
- [40] Mougang, F.J., and Zanga, A.D. Stage deformation sur le développement de la pisciculture. *The Egyptian International Centre for Agriculture*. 29p, 2015.
- [41] V. N'douba, and A Lambert, "Deux Monogènes nouveaux parasites branchiaux de *Clarias ebriensis* Pellegrin, 1920 (Siluriforme, Clariidae) en Côte d'Ivoire", *Zoosystema* vol. 23, pp 411-416, 2001.

- [42] J. Nack, C. F. Bilong Bilong and L. Euzet, "Monogènes parasites de Clariidae (Teleostei, Siluriformes) au Cameroun: I Description de deux nouvelles espèces du genre *Gyrodactylus* dans le bassin du Nyong", *Parasite*. vol. 12, pp 213-220, 2005.
- [43] J. Nack, J. Massende, and D. F. Messu Mandeng, "Distribution spatiale de deux monogènes du genre *Protoancylodiscoides* Paperna, 1969 (Dactylogyridea, Ancyrocephalidae) parasites branchiaux de *Chrysichthys auratus* (Geoffroy Saint-Hilaire, 1808) (Siluriformes, Clariidae) dans le Lac Ossa (Littoral-Cameroun)", *Journal of Applied. Biosciences*.vol. 121, pp 12157-12167, 2018.
- [44] D. A. Nomwine, R. Ouédraogo, and A. Ouéda, "Relation poids-longueur et facteur de condition de *Clarias anguillaris* et *Sarotherodon galilaeus* pêchées dans le Lac Bam et le réservoir de la Komienga au Burkina Faso", *International Journal of. Biological and Chemical Sciences*. vol. 12, no. 4, pp 1601-1610, 2018.
- [45] Obiekezie, A.I. *The principal pathogens and diseases of cultured fishes in Nigeria*. In. Koop (Ed). Aquaculture in Africa. A workshop on research in Aquaculture Harare , Zimbabwe 23th-27th January 1991; IFS Publication: pp 197-207, 1991.
- [46] A. I. Obiekezie, H. Muller and K. Anders, "Diseases of the african estuarine catfish *Chrysichthys nigrodigitatus* (Lacépède) from the Cross River estuary, Nigeria", *Journal of fish Biology*. vol. 32, pp 207-221. 1988.
- [47] Olivry, J. C. Fleuves et rivières du Cameroun. Monographies Hydrologiques ORSTOM. Paris, MESRES-ORSTOM., 781p. 1986.
- [48] I. Paperna, "Dynamics of *Dactylogyrus vastator* Nybelin (Monogenea) populations on the gills of carp fry in fish ponds", *Bamidgeh, Bulletin of Fish Culture in Israel*, vol.15 pp 31-50, 1963.
- [49] Poulin, R. Evolutionary Ecology of parasites from individuals to communities. Ed Chapman de Hall. 212p, 1998.
- [50] A. Soleng and T. A. Bakke, "The suscepibility of grawling (*Thymallus thymallus*) to the experimental infection with the monogenean *Gyrodactylus salaris*", *International Journal for Parasitology*, vol. 31,pp 793-797, 2002.
- [51] M. L. J. Stiasny, G.G. Teugels and C. D. Hopkins, *Poissons d'eaux et saumâtres de basse Guinée, ouest de l'Afrique centrale*. Vol 1. IRD, MnHn, MRAC, Paris, 800 p, 2007.
- [52] Suchel J. B., Les climats du Cameroun. (Thèse de Doctorat d'Etat). Université de Bordeaux III. 1186p. 1987.
- [53] G.G. Teugels, "A systematic revision of the African species of the genus *Clarias* (Pisces, Clariidae).Annales du Musée Royal d'Afrique Centrale", *Sciences Zoologiques*. Vol. 247, pp 1-199, 1986.
- [54] G. G. Teugels and D. Adriaens, "Taxonomy and phylogeny of Clariidae: an overview, p. 465-487. In: Arratia G, Kapoor B, Chardon M, Diego R (Eds.) Catfishes. Enfield, Science publisherInc.Wirrmann D. (1992) Le Lac Ossa: une monographie préliminaire", *Revue Géographique du Cameroun*, vol. 11, pp 27-38, 2003.
- [55] R. C. Tinsley and J. A Jackson, "Host factors limiting monogeneans infection. A case study", *International Journal for Parasitology*. vol.32, pp 353-365, 2002.
- [56] T.C.T.iogue, D Nguenga, M. Tomedi Eyango, and J. Tchoumboue, "Quelques performances reproductives et taux de survie de deux souches du poisson chat africain *Clarias gariepinus* Burchell, 1822) et de leurs croisés à Koupa-Matapit", *International Journal of Biological and Chemical Sciences*, vol. 2, no. 4, :pp 469-477, 2008.
- [57] H. C Tozato, S. P. Carlos and S. P. Garça, "Influência do parasitismo na condição de *Corydoras aeneus* (Gill,1858) (Osteichthyes: Siluriformes) da bacia do ribeirão do Feijão, São", . *Revista científica eletrônica de Medicina Veterinária*, vol. 16, pp 1-15, 2011.
- [58] E. Unlu and K Balci, "Observation on the reproduction of *Leuciscus cephalus orientalis* (Cyprinidae in Savour Stream (Turkey)", *Cybium*. vol. 17, no. 3, pp 241-240, 1993.
- [59] E. T. Valtonen, J. C. Holmes, and M. Koskivaara, "Eutrophication, pollution and fragmentation : effects on parasite communities in roach (*Rutilus rutilus*) and perch (*Perca fluviatilis*) in four lakes in the Central Finland", *Canadian Journal of Fisheries and Aquatic Sciences*. vol. 54, :pp 572-585, 1997.
- [60] P. Zango, M. Tabi Eyango Tomedi, T. Ewoukem Efole, C. Tekounegning Tiogue, D. Nguenga, S. M. Kamanke Kamanke, O. Mikolasek and J. Tchoumboue, "Performances de reproduction du poisson chat endogène du Cameroun *Clarias jaensis* (Boulenger, 1909) en milieu contrôlé", *Intertional Journal Biological and Chemical Sciences*, vol. 10, no. 2, pp 533-542, 2016.
- [61] P. Zango, M. Tabi Eyango Tomedi, C. Tekounegning Tiogue, T. Ewoukem Efole, V. Pouomegne, O. Mikolaseck and J. Tchoumboue, "Performances comparée de survie et croissance de *Oreochromis niloticus* associé à *Clarias jaensis* et *Clarias gariepinus*", . *Cameroon Journal of Experimental Biology*. vol. 11, no. 1, pp 1-8, 2017.